



Sample Grant Text
Perelman School of Medicine

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Note: The text provided in this document is sample language; please review your RFA carefully to ensure that you provide what is requested, and edit as this text as appropriate.

About this Resource

The information provided in this document is offered to assist investigators in the preparation of grant submissions. Content featured is a compilation of information provided by UPenn faculty, who were generous enough to share their sample text.

Boilerplate text may be copied into grants and used as a foundation to create individualized proposals. Please note that the information provided is not exhaustive and may not be applicable to specific funders or grant guidelines. Please review both the grant guidelines and the boilerplate text carefully, and use only what is appropriate.

The provided text will evolve over time and undergo periodic updates. In addition to the sample text, please refer to our [PSOM Grant Support](#) for links to additional data resources. Please note that some grant sections require inputting current facts; investigators are encouraged to tailor these statements for your specific proposal as appropriate.

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AUTHENTICATION OF KEY RESOURCES

General: The proposed application includes study of (clinical trial or animal studies). All findings related to these studies will be recorded in laboratory notebooks that are maintained in a secure location in the laboratory and/or stored in electronic databases that are saved on password-protected computers. All tissues collected from animal or human subjects are identified only by study id for the purpose of assays by collaborating investigators. Thus, collaborating investigators are blinded to treatment groups. Study ids are HIPPA compliant and all human patient data is de-identified in laboratory records. Publications will include sufficient experimental detail to enable independent reproduction of methods. Raw data that is used to calculate summary statistics in publications will be provided in supplementary data as required by the journal or otherwise be available by request.

Antibodies: Antibodies to be used in this study are to be purchased through commercial vendors that include XXX and XXX. Every attempt will be made to consistently obtain antibodies of the same lot number for the duration of a study. Antibodies will be reconstituted (if needed), stored per manufacturer's instructions and used prior to the expiration date. Validation of new batches or lots of antibodies for signaling molecules will be performed via western blotting with comparisons to existing lots and confirmation of the protein band of interest in cells known to be positive for the protein. When possible, RNAi oligonucleotides will be employed to ensure that a band of appropriate molecular weight is no longer detected after knockdown. Antibodies to be used for immunohistochemistry will be tested in banked tissues previously determined to be positive for the marker of interest, with the specificity of staining confirmed via appropriate blocking peptides and/or competitive inhibition with a second antibody. Additional controls include replacement of the primary antibody with a nonimmune Ig (generated in the same animal species as was the primary antibody) and exclusion of the primary antibody in the case of staining protocols that use a secondary antibody. Antibodies for flow cytometry will be similarly validated using isotype controls and investigation of known positive/negative cell types.

Verification of Cell Lines: Whenever possible cells lines will be obtained from commercial cell banks (e.g., ATCC). In the case of cell lines obtained from other laboratories, the lines will be subject to authenticity and mycoplasma testing (e.g., by IDEXX Bioresearch) before use in experiments. Any positive results will be reported to the donating laboratory. If cells are positive for mycoplasma, they will be treated for contamination using an accepted antimycoplasma reagent (e.g., by Lonza) and retested prior to use. All cells lines in use in the laboratory are subject to authenticity on an annual basis. Cells are monitored for contamination (e.g., fungus or yeast) on an ongoing basis by visualization under a microscope. Cells maintained in vitro are identified by serial passage numbers with designated passage numbers used for in vitro and in vivo studies.

Transgenic Animals: XXXX transgenic mice will be procured from XXXXX. The genotypes will be confirmed using validated primers. Validation of the genotypes will be done in all progenies.

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Drugs/Chemicals: *Plasmids:* All plasmids (expression and lentiviral vectors) obtained from other laboratories or generated in our laboratory will be subjected to sequencing to confirm their identity prior to any use. *Specialty Chemicals:* Chemicals for these studies will be purchased from reputable commercial vendors, such as Sigma-Aldrich. All reagents will be stored as recommended and utilized by their expiration date. All compounds that are frozen will be stored in batches at -20°C or -80°C as appropriate, and subjected only to a single freeze-thaw cycle. All studies will include appropriate controls to ensure the expected biological activity. **XX Add information here on the source and storage of any specific drugs to be used in the proposed studies XXX**